

Mild Rh(III)-Catalyzed C7-Allylation of Indolines with Allylic Carbonates

Jihye Park,[†] Neeraj Kumar Mishra,[†] Satyasheel Sharma,[†] Sangil Han,[†] Youngmi Shin,[†] Taejoo Jeong,[†] Joa Sub Oh,[‡] Jong Hwan Kwak,[†] Young Hoon Jung,[†] and In Su Kim^{*,†}

[†]School of Pharmacy, Sungkyunkwan University, Suwon 440-746, Republic of Korea

[‡]College of Pharmacy, Dankook University, Cheonan 330-714, Republic of Korea

S Supporting Information



ABSTRACT: The rhodium(III)-catalyzed direct allylation of indolines with allylic carbonates at room temperature is described. These transformations provide the facile and efficient construction of C7-allylated indolic scaffold.

INTRODUCTION

Transition-metal-catalyzed C–H bond functionalization has emerged as a powerful tool due to its remarkable potential for step economy and environmental sustainability.¹ With the development of catalytic C–H bond functionalization, it has become the most straightforward protocol leading to allylated molecules. For example, Oi and Inoue first reported the ruthenium-catalyzed direct allylation of 2-phenylpyridines with allylic acetates affording a regioisomeric mixture of olefins.² Bergman and Ellman described a single example for the Rh(III)-catalyzed allylation of a ketoxime with allyl acetate in the presence of copper oxidant to deliver a terminal alkene.³ Glorius disclosed a beautiful protocol on the Rh(III)-catalyzed terminal allylation of benzamides and indoles with allylic carbonates.⁴ In sharp contrast, Loh demonstrated the Rh(III)-catalyzed internal allylation of benzamides with allylic acetates to provide conjugated olefins.⁵ Friedel–Crafts-type C–H allylations of aromatic compounds with allylic carbonates under ruthenium catalysis were also demonstrated.⁶ Very recently, direct C–H allylations of electron-deficient poly-fluoroarenes and heterocycles under copper and palladium catalysis were reported.⁷ Moreover, Krische,⁸ Ma,⁹ and Cramer¹⁰ independently described the Ir- or Rh-catalyzed allylation reactions of benzamides using allenes as allyl sources. Interestingly, Glorius has recently reported the selective and efficient Co(III)-catalyzed C–H allylation of indoles with allyl carbonates.¹¹ In addition, Wang demonstrated the Rh(III)-catalyzed direct C–H allylation reaction with 4-vinyl-1,3-dioxolan-2-ones to afford allylic alcohols.¹²

The indoles and indolines are ubiquitous structural motifs found in a large number of natural products with diverse and important biological activities.¹³ In particular, the allylated indole or indoline alkaloids are widely distributed in terrestrial and marine organisms, especially in the genera *Penicillium* and

Aspergillus of ascomycota, and display broad structural diversity.¹⁴ The prevalence of allylated indoles and indolines in bioactive natural products and the versatility of olefin transformations have led to the development of many useful methods for their preparation.^{15,16} Notably, the C7-allylated indoles and indolines are known as pivotal heterocyclic compounds found in a number of bioactive synthetic molecules and natural products, as shown in Figure 1.¹⁷

Recently, the directing group-assisted catalytic C7-functionalizations of indolines with various coupling partners were demonstrated (Scheme 1). For example, acylation,¹⁸ arylation,¹⁹ olefination,²⁰ alkylation,²¹ and alkynylation²² of indo-

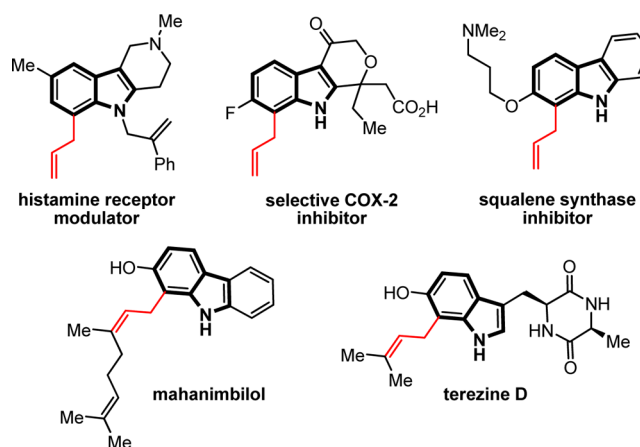


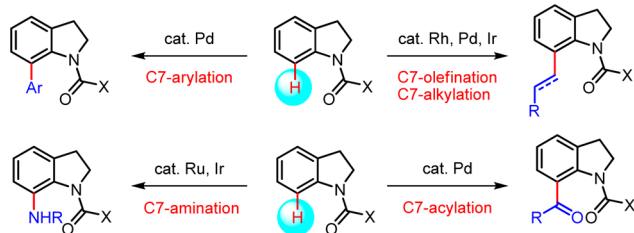
Figure 1. Selected examples for synthetic or natural C7-allylated products.

Received: December 3, 2014

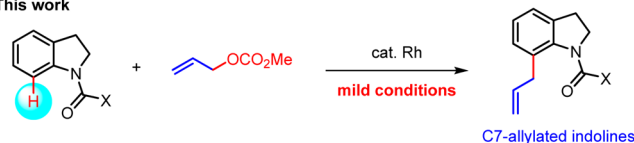
Published: January 13, 2015

Scheme 1. Catalytic C7-Functionalization of Indolines

Previous works



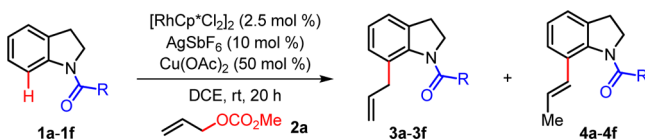
This work



lines at the C-7 position were described under ruthenium, palladium, rhodium, iron, and iridium catalysis. In addition, Ru(II)- or Ir(III)-catalyzed C7-aminations of indolines with organic azides were reported by Zhu, Chang, and Zhou/Li, respectively.²³ Inspired by our recent study on the decarboxylative acylation of indolines at the C7-position²⁴ and in continuation of the catalytic C–H allylation of aromatic and α,β -unsaturated carboxamides,²⁵ we herein disclose the rhodium-catalyzed direct allylation and crotylation of indolines with allylic carbonates via C–H bond activation.

RESULTS AND DISCUSSION

Our study was initiated by examining the coupling of *N*-acetyl indoline (**1a**) and allyl methyl carbonate (**2a**) under rhodium catalysis (Table 1). To our delight, the cationic rhodium

Table 1. Screening of *N*-Protection Groups^a

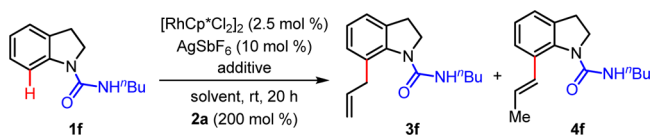
entry	R	substrate	yield (%) ^b	ratio (3:4) ^c
1	Me	1a	55	1.9:1
2	<i>t</i> -Bu	1b	37	1.4:1
3	NMe ₂	1c	35	6.9:1
4	pyrrolidinyl	1d	12	3.5:1
5	NHPh	1e	19	10.9:1
6	NH ^{<i>t</i>} Bu	1f	80	9.5:1

^aReaction conditions: **1a–1f** (0.3 mmol), **2a** (0.6 mmol), [RhCp*Cl₂]₂ (2.5 mol %), AgSbF₆ (10 mol %), Cu(OAc)₂ (50 mol %), DCE (1 mL) under air at room temperature for 20 h in reaction tubes. ^bIsolated yield by flash column chromatography. ^cRegioisomeric ratio between terminal and internal olefins determined by ¹H NMR analysis of crude reaction mixture.

complex, derived from [Cp*^{*Rh*}Cl₂]₂ and AgSbF₆, was found to catalyze the coupling of **1a** and **2a** in the presence of Cu(OAc)₂ as an additive in dichloroethane (DCE) at room temperature for 20 h to provide the C7-allylated product **3a** with a regioisomeric mixture of 1.9:1 ratio in 55% yield (Table 1, entry 1). Similarly, pivaloyl directing group **1b** displayed low level of both reactivity and selectivity (Table 1, entry 2). However, *N,N*-dimethylcarbamoyl indoline (**1c**) gave terminal olefin **3c** with good level of regioselectivity (6.9:1), albeit in low yield

(Table 1, entry 3). Thus, we focused on the screening of other carbamoyl directing groups. Interestingly, *N*-butylindoline-1-carboxamide (**1f**), derived from indoline and *n*-butyl isocyanate, was found to be far more effective in this coupling reaction to afford our desired product **3f** with high terminal selectivity (9.5:1) in 80% yield (Table 1, entry 6).

With the optimal directing group of indolines in hand, we further optimized the reaction conditions to afford increased terminal selectivity (Table 2). Screening of solvents under

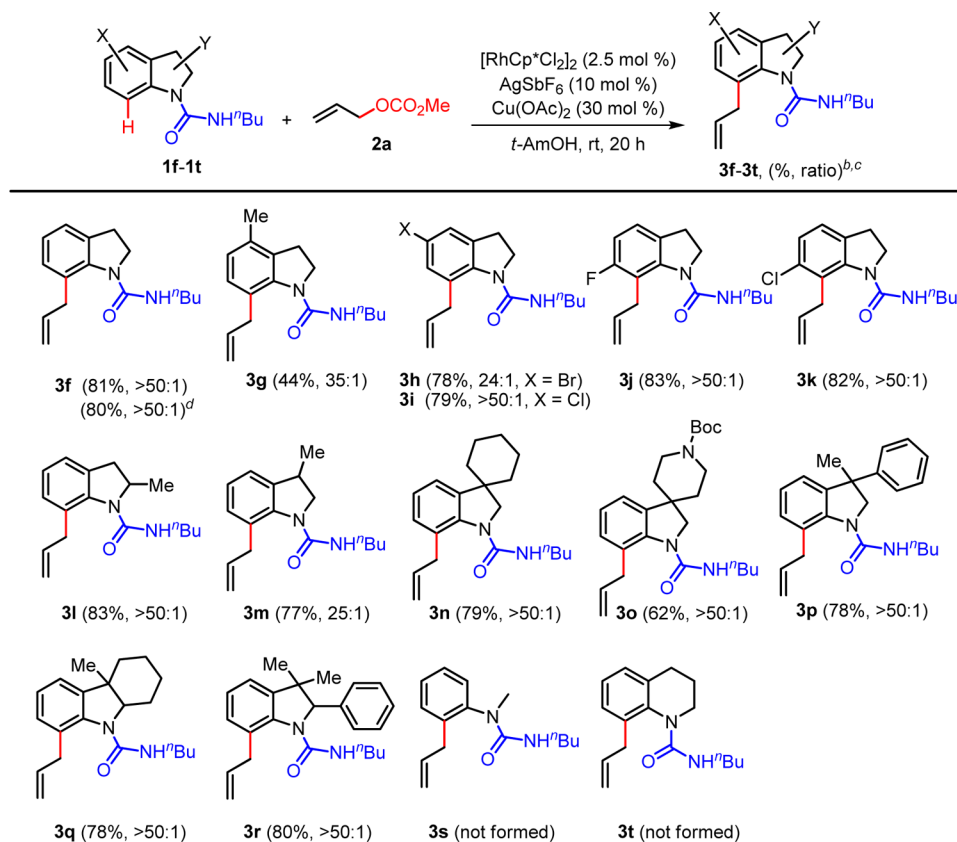
Table 2. Selected Optimization of the Reaction Conditions^a

entry	additive (mol %)	solvent	yield (%) ^b	ratio (3f:4f) ^c
1	Cu(OAc) ₂ (50)	DCE	80	9.5:1
2	Cu(OAc) ₂ (50)	DMF	39	5.7:1
3	Cu(OAc) ₂ (50)	toluene	29	7.5:1
4	Cu(OAc) ₂ (50)	MeCN	5	1.0:1
5	Cu(OAc) ₂ (50)	THF	74	8.9:1
6	Cu(OAc) ₂ (50)	<i>t</i> -AmOH	84	>50:1
7	Cu(OAc) ₂ ·H ₂ O (50)	<i>t</i> -AmOH	66	7.1:1
8	PivOH (50)	<i>t</i> -AmOH	5	>50:1
9	AgOAc (50)	<i>t</i> -AmOH	57	6.3:1
10	Cu(OAc) ₂ (30)	<i>t</i> -AmOH	81	>50:1
11		<i>t</i> -AmOH	NR	trace
12 ^d	Cu(OAc) ₂ (30)	<i>t</i> -AmOH	NR	0

^aReaction conditions: **1f** (0.3 mmol), **2a** (0.6 mmol), [RhCp*Cl₂]₂ (2.5 mol %), AgSbF₆ (10 mol %), additive (quantity noted), solvent (1 mL) under air at room temperature for 20 h in reaction tubes. ^bIsolated yield by flash column chromatography. ^cRegioisomeric ratio between terminal and internal olefins determined by ¹H NMR analysis of crude reaction mixture. ^dIn the absence of AgSbF₆.

otherwise identical conditions revealed that *tert*-amyl alcohol (*t*-AmOH) was found to be the most effective solvent in this coupling reaction to afford terminal olefination product **3f** in high yield (84%) with excellent level of regioselectivity (>50:1), whereas other solvents such as DCE, DMF, toluene, MeCN, and THF were less effective (Table 2, entries 1–6). Further study revealed that Cu(OAc)₂ additive is unique in its ability to facilitate high levels of conversion and selectivity (Table 2, entries 7–9). In addition, decreasing amount of Cu(OAc)₂ to 30 mol % provided a comparable yield (81%) and selectivity (>50:1) (Table 2, entry 10), but no formation of **3f** was observed when either Cu(OAc)₂ or AgSbF₆ were excluded (Table 2, entries 11 and 12).

To evaluate the scope and limitation of this process, the optimal reaction conditions were applied to a range of *N*-butyl carbamoyl indolines **1g–1t** (Table 3). In all cases, good to excellent yields and high terminal selectivity of the desired aryl C–H allylation adducts were obtained. Exceptionally, indoline **1g** with electron-rich moiety at the C4-position exhibited slightly decreased reactivity. It should be noted that C6-substituted indolines **1j** and **1k** were tolerated under current reaction conditions to afford the corresponding products in high yields and excellent level of regioselectivity. In addition, this reaction was highly compatible with C2-substituted indolines **1l**, **1q**, and **1r**. To our surprise, this reaction is only restricted to indolines. For example, *N*-methyl aniline **1s** and tetrahydroquinoline **1t** containing a *N*-butylcarbamoyl directing

Table 3. Scope of Indolines^a

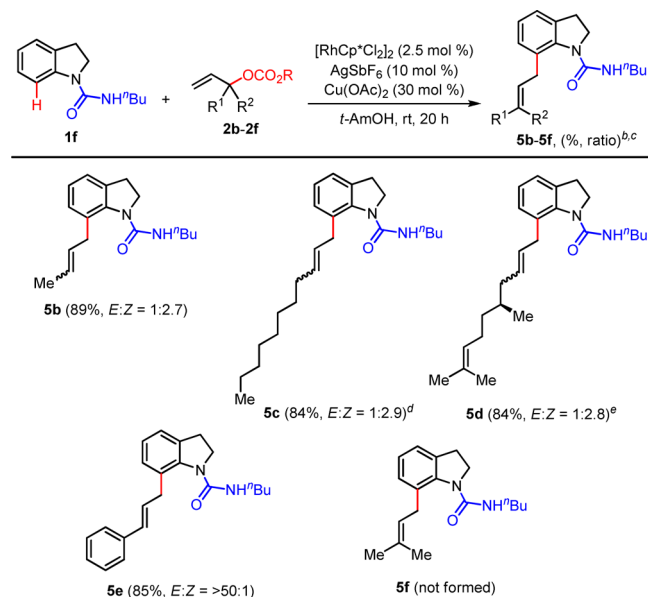
^aReaction conditions: **1f-1t** (0.3 mmol), **2a** (0.6 mmol), $[\text{RhCp}^*\text{Cl}_2]_2$ (2.5 mol %), AgSbF_6 (10 mol %), $\text{Cu}(\text{OAc})_2$ (30 mol %), *t*-AmOH (1 mL) under air at room temperature for 20 h in reaction tubes. ^bIsolated yield by flash column chromatography. ^cRegioisomeric ratio between terminal and internal olefins determined by ¹H NMR analysis of crude reaction mixture. ^dScale-up experiment (**1f**, 3 mmol).

group failed to deliver the corresponding products under standard reaction conditions.

To further explore the scope and limitation of this transformation, various allylic carbonates **2b-2f** were screened to couple with **1f**, as shown in Table 4. The monosubstituted allyl carbonates **2b-2d** were smoothly coupled with **1f** to give the (*Z*)-crotylation products **5b-5d** as major isomers in high yields. In sharp contrast, α -phenyl-substituted allyl carbonate **2e** underwent completely *trans*-selectivity with **1f**. Notably, these reactions proceeded readily with complete γ -selectivity in case of branched allylic carbonates, and no migration of double bond on the products was observed. However, linear crotyl carbonates and β -substituted allylic carbonates did not deliver the corresponding coupling products, presumably due to the increased steric hindrance of electrophilic allylic carbonates, preventing formal S_{N}^2 -type reactions with rhodacycle intermediate.²⁶ In addition, α,α -disubstituted allylic carbonate **2f** was found to be unreactive under the optimal reaction conditions.

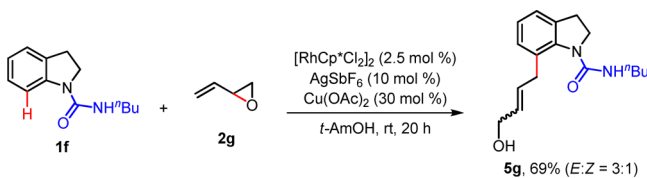
To our delight, 2-vinylloxirane **2g** was also coupled with indoline **1f** to afford a mixture of allylic alcohol **5g** with 3:1 *E/Z* ratio in 69% yield as a result of olefin insertion and epoxide ring-opening, which is in agreement with the formal S_{N}^2 -type reaction mechanism (Scheme 2).²⁷

To demonstrate the synthetic utility of C7-allylated indolines, various transformations of the allylated product **3f** were conducted (Scheme 3). Intermolecular olefin metathesis between **3f** and ethyl acrylate gave α,β -unsaturated ester **6a**, which subsequently underwent aza-Michael reaction to furnish

Table 4. Scope of Allylic Carbonates^a

^aReaction conditions: **1f** (0.3 mmol), **2b-2f** (0.6 mmol), $[\text{RhCp}^*\text{Cl}_2]_2$ (2.5 mol %), AgSbF_6 (10 mol %), $\text{Cu}(\text{OAc})_2$ (30 mol %), *t*-AmOH (1 mL) under air at room temperature for 20 h in reaction tubes. All cases provided the regioisomeric ratio of above 50:1 between terminal and internal olefins. ^bIsolated yield by flash column chromatography. ^cDiastereomeric ratio between *trans*- and *cis*-olefins determined by ¹H NMR analysis. ^dTHF was used as a solvent. ^e40 h.

Scheme 2. Indoline Allylation with 2-Vinyloxirane



tricyclic compound **6b** in 45% yield. Interestingly, while carrying out the deprotection of *N*-butylcarbamoyl group of **3f** under standard basic conditions, we observed the deprotection as well as olefin migration of **3f** to provide free-(NH)-indoline **6c** in high yield. Subsequently this deprotected product **6c** under transfer hydrogenation conditions provided smoothly C7-olefinated indole **6d** in high yield.

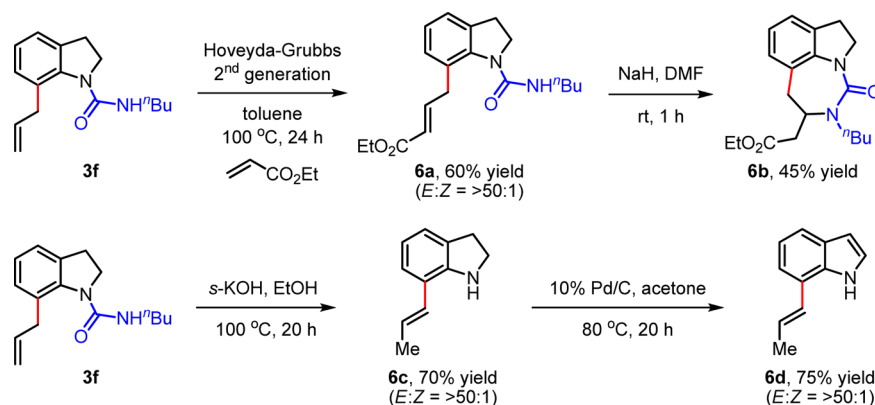
To gain mechanistic insight, the following experiments were performed (Scheme 4). A hydrogen–deuterium exchange using MeOD indicated that the cleavage of the C–H bond at the indoline C7-position was a reversible metalation–proto-(deutero)demetalation process (Scheme 4a). In the presence of allyl methyl carbonate (**2a**) and MeOD, no deuterium incorporation of product **3a** and significant deuteriation (71% D) of recovered starting material was observed (Scheme 4b). Next, two parallel reactions of **1f** and **deuterio-1f** with **2a** under standard reaction conditions resulted in the kinetic isotope effect (k_H/k_D) of 2.89, thus indicating that C–H cleavage might be involved in the rate-limiting step (Scheme 4c).²⁸

A proposed reaction mechanism was depicted in Scheme 5. Coordination of the carbonyl group to cationic Rh(III) catalyst and subsequent C–H cleavage delivers the cyclorhodated species **I**, which on migratory insertion of a double bond into Rh–C bond affords an eight-membered Rh(III) intermediate **II**. Further, β -oxygen elimination provides the product **3f** and regenerates a Rh(III) catalyst. Alternatively, coordination of allyl methyl carbonate to the cyclorhodated species **I** followed by nucleophilic substitution can not be ruled out in the catalytic cycle to afford 7-allylated product **3f**.²⁹

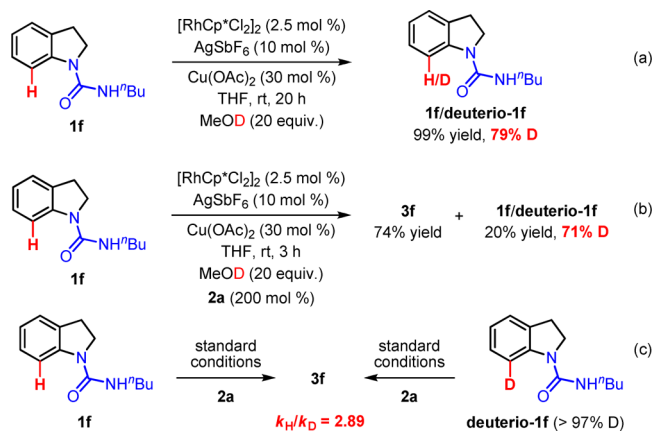
CONCLUSION

We have disclosed a highly selective C7-allylation of indolines with allylic carbonates under rhodium catalysis. These transformations have been applied to a wide range of substrates, and typically proceed with excellent levels of chemoselectivity as well as with high functional group tolerance.

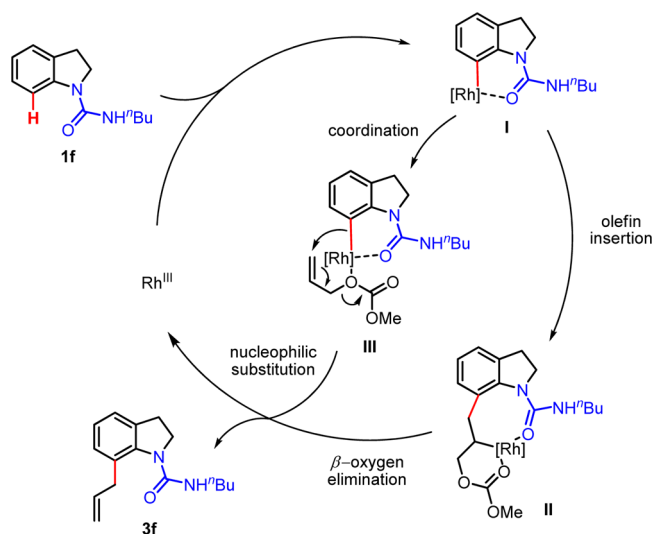
Scheme 3. Synthetic Transformation of C7-Allylated Indolines



Scheme 4. Mechanistic Investigations



Scheme 5. Proposed Reaction Mechanism



EXPERIMENTAL SECTION

General Procedure for the Synthesis of *N*-Acyl or *N*-Carbamoyl Indolines (1a–1e**).** To a stirred solution of indoline (1.00 g, 8.4 mmol) and triethylamine (3.5 mL, 25.2 mmol) in CH_2Cl_2 (10 mL) was added dropwise a solution of *N*-acyl or *N*-carbamoyl chloride (12.6 mmol) in CH_2Cl_2 (7 mL) at 0 °C. The reaction mixture was stirred at this temperature for 15 min and further stirred at room temperature for 3 h. The resulting mixture was partitioned between CH_2Cl_2 and H_2O . The organic layer was dried over MgSO_4 and

concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc) to give the corresponding products **1a–1e**.

1-(Indolin-1-yl)ethanone (1a).^{20b} 1.21 g (89%); white solid; mp = 102–104 °C; ¹H NMR (700 MHz, CDCl₃) δ 8.18 (d, *J* = 8.0 Hz, 1H), 7.18–7.14 (m, 2H), 6.98 (t, *J* = 7.4 Hz, 1H), 4.03 (t, *J* = 8.4 Hz, 2H), 3.18 (t, *J* = 8.4 Hz, 2H), 2.20 (s, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 168.7, 142.9, 131.0, 127.5, 124.5, 123.6, 117.0, 48.7, 28.0, 24.3.

1-(Indolin-1-yl)-2,2-dimethylpropan-1-one (1b).^{23a} 1.33 g (78%); light brown solid; mp = 54–58 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.23 (d, *J* = 8.5 Hz, 1H), 7.19–7.17 (m, 2H), 7.02 (t, *J* = 7.0 Hz, 1H), 4.23 (t, *J* = 8.1 Hz, 2H), 3.14 (t, *J* = 8.1 Hz, 2H), 1.38 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 176.5, 144.7, 130.7, 127.3, 124.2, 123.6, 118.4, 49.4, 40.2, 29.3, 27.7.

***N,N*-Dimethylindoline-1-carboxamide (1c).**^{20b} 1.32 g (83%); light brown solid; mp = 57–62 °C; ¹H NMR (700 MHz, CDCl₃) δ 7.13 (d, *J* = 7.2 Hz, 1H), 7.10 (t, *J* = 7.9 Hz, 1H), 6.90 (d, *J* = 7.9 Hz, 1H), 6.85 (t, *J* = 7.3 Hz, 1H), 3.87 (t, *J* = 8.2 Hz, 2H), 3.00 (t, *J* = 8.2 Hz, 2H), 2.91 (s, 6H); ¹³C NMR (175 MHz, CDCl₃) δ 160.3, 144.3, 131.4, 120.7, 124.8, 121.3, 113.3, 50.3, 38.1, 28.1.

Indolin-1-yl(pyrrolidin-1-yl)methanone (1d). 1.25 g (69%); light yellow solid; mp = 76–85 °C; ¹H NMR (700 MHz, CDCl₃) δ 7.08–7.03 (m, 3H), 6.79 (t, *J* = 7.2 Hz, 1H), 3.85 (t, *J* = 8.3 Hz, 2H), 3.39–3.37 (m, 2H), 3.29–3.26 (m, 2H), 2.97 (t, *J* = 8.4 Hz, 2H), 1.82–1.80 (m, 2H), 1.75–1.72 (m, 2H); ¹³C NMR (175 MHz, CDCl₃) δ 158.5, 144.3, 131.1, 126.9, 124.5, 121.3, 114.7, 49.9, 48.3, 47.8, 38.2, 28.5, 25.4; IR (KBr) ν 2969, 2873, 1637, 1600, 1478, 1384, 1259, 1204, 1017, 917, 871, 730 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₃H₁₆N₂O [M]⁺ 216.1263, found 216.1262.

***N*-Phenylindoline-1-carboxamide (1e).** 1.40 g (70%); white solid; mp = 109–111 °C; ¹H NMR (700 MHz, CDCl₃) δ 7.87 (d, *J* = 7.9 Hz, 1H), 7.41 (d, *J* = 7.7 Hz, 2H), 7.28 (br s, 2H), 7.18–7.14 (m, 2H), 7.04 (br s, 1H), 6.93 (t, *J* = 6.9 Hz, 1H), 6.53 (br s, 1H), 4.01–3.94 (m, 2H), 3.16–3.13 (m, 2H); ¹³C NMR (175 MHz, CDCl₃) δ 152.4, 144.2, 138.3, 130.7, 129.0, 127.7, 124.8, 123.6, 122.3, 120.3, 115.0, 47.5, 27.9; IR (KBr) ν 3316, 3048, 2884, 1652, 1523, 1478, 1438, 1342, 1237, 1171, 1141, 1024, 732 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₅H₁₄N₂O [M]⁺ 238.1106, found 238.1107.

General Procedure for the Synthesis of *N*-Butylindoline-1-carboxamides (1f–1r). To a stirred solution of indoline (1.00 g, 8.4 mmol) in CH₂Cl₂ (25 mL) were added triethylamine (3.5 mL, 25.2 mmol) and *n*-butyl isocyanate (1.25 g, 12.6 mmol) at 0 °C. The reaction mixture was then stirred at room temperature for 3 h. The reaction mixture was washed with H₂O and extracted with EtOAc. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc) to give the corresponding products **1f–1r**.

***N*-Butylindoline-1-carboxamide (1f).** 1.48 g (81%); white solid; mp = 72–74 °C; ¹H NMR (700 MHz, CDCl₃) δ 7.84 (d, *J* = 8.0 Hz, 1H), 7.13–7.09 (m, 2H), 6.86 (t, *J* = 7.4 Hz, 1H), 4.60 (br s, 1H), 3.85 (t, *J* = 8.4 Hz, 2H), 3.30–3.28 (m, 2H), 3.12 (t, *J* = 8.6 Hz, 2H), 1.54–1.50 (m, 2H), 1.38–1.33 (m, 2H), 0.92 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 155.1, 143.8, 130.2, 127.6, 124.5, 121.6, 114.6, 47.0, 40.3, 32.3, 27.8, 20.1, 13.8; IR (KBr) ν 3344, 2956, 2928, 2861, 1643, 1520, 1480, 1461, 1386, 1339, 1292, 1264, 1153, 1088, 1022, 930, 748 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₃H₁₈N₂O [M]⁺ 218.1419, found 218.1420.

***N*-Butyl-4-methylindoline-1-carboxamide (1g).** 1.42 g (73%); yellow sticky solid; ¹H NMR (700 MHz, CDCl₃) δ 7.68 (d, *J* = 8.8 Hz, 1H), 6.94–6.92 (m, 2H), 4.52 (br s, 1H), 3.85 (t, *J* = 8.3 Hz, 2H), 3.31–3.28 (m, 2H), 3.09 (t, *J* = 8.5 Hz, 2H), 2.25 (s, 3H), 1.54–1.50 (m, 2H), 1.39–1.33 (m, 2H), 0.92 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 155.3, 141.5, 131.3, 130.6, 128.1, 125.5, 114.4, 47.3, 40.4, 32.5, 28.0, 21.0, 20.3, 14.0; IR (KBr) ν 3338, 2957, 2928, 2861, 1644, 1523, 1487, 1384, 1332, 1263, 1160, 1133, 817, 733 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₄H₂₀N₂O [M]⁺ 232.1576, found 232.1577.

5-Bromo-*N*-butylindoline-1-carboxamide (1h). 1.74 g (70%); light yellow solid; mp = 159–162 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.82 (d, *J* = 7.3 Hz, 1H), 7.27–7.24 (m, 2H), 4.52 (br s, 1H), 3.90 (t, *J* =

7.8 Hz, 2H), 3.33 (s, 2H), 3.17 (t, *J* = 7.9 Hz, 2H), 1.61–1.51 (m, 2H), 1.45–1.33 (m, 2H), 0.96 (t, *J* = 6.6 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 154.8, 143.0, 132.2, 130.4, 127.4, 116.1, 113.8, 47.0, 40.2, 32.2, 27.5, 20.1, 13.8; IR (KBr) ν 3343, 2956, 2928, 2871, 2176, 1645, 1519, 1468, 1327, 1298, 1253, 1154, 1090, 817, 734 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₃H₁₇BrN₂O [M]⁺ 296.0524, found 296.0522.

***N*-Butyl-5-chloroindoline-1-carboxamide (1i).** 1.44 g (68%); light yellow solid; mp = 87–91 °C; ¹H NMR (300 MHz, CDCl₃) δ 8.04 (d, *J* = 8.1 Hz, 1H), 7.43 (s, 1H), 7.29–7.26 (m, 1H), 4.68 (br s, 1H), 4.07 (t, *J* = 8.9 Hz, 2H), 3.49–3.47 (m, 2H), 3.33 (t, *J* = 8.4 Hz, 2H), 1.77–1.68 (m, 2H), 1.62–1.50 (m, 2H), 1.15–1.10 (m, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 154.9, 142.5, 131.8, 127.4, 126.4, 124.5, 115.6, 47.1, 40.2, 32.2, 27.6, 20.1, 13.8; IR (KBr) ν 3340, 2957, 2930, 2872, 1645, 1519, 1471, 1328, 1252, 1154, 1091, 874, 819, 733 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₃H₁₇ClN₂O [M]⁺ 252.1029, found 252.1034.

***N*-Butyl-6-fluoroindoline-1-carboxamide (1j).** 1.29 g (65%); white solid; mp = 114–116 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.68 (d, *J* = 8.6 Hz, 1H), 7.00 (t, *J* = 7.9 Hz, 1H), 6.59–6.53 (m, 1H), 4.53 (br s, 1H), 3.91 (t, *J* = 9.0 Hz, 2H), 3.34–3.28 (m, 2H), 3.12 (t, *J* = 8.5 Hz, 2H), 1.59–1.50 (m, 2H), 1.44–1.32 (m, 2H), 0.94 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 162.0 (d, *J*_{C-F} = 239.5 Hz), 154.8, 145.1 (d, *J*_{C-F} = 12.0 Hz), 125.1 (d, *J*_{C-F} = 2.5 Hz), 124.5 (d, *J*_{C-F} = 10.3 Hz), 107.8 (d, *J*_{C-F} = 22.5 Hz), 103.0 (d, *J*_{C-F} = 29.7 Hz), 47.8, 40.2, 32.2, 27.1, 20.1, 13.8; IR (KBr) ν 3338, 2954, 2931, 2870, 1643, 1530, 1490, 1390, 1300, 1263, 1160, 1079, 856, 786, 738 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₃H₁₇FN₂O [M]⁺ 236.1325, found 236.1329.

***N*-Butyl-6-chloroindoline-1-carboxamide (1k).** 1.29 g (61%); light yellow solid; mp = 83–85 °C; ¹H NMR (700 MHz, CDCl₃) δ 7.96 (s, 1H), 7.00 (d, *J* = 7.5 Hz, 1H), 6.84 (d, *J* = 7.6 Hz, 1H), 4.51 (br s, 1H), 3.89 (t, *J* = 8.4 Hz, 2H), 3.31 (t, *J* = 6.8 Hz, 2H), 3.12 (t, *J* = 8.4 Hz, 2H), 1.60–1.50 (m, 2H), 1.44–1.32 (m, 2H), 0.94 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 154.8, 145.0, 133.3, 124.9, 121.5, 115.2, 47.5, 40.3, 32.3, 27.4, 20.1, 13.8; IR (KBr) ν 3340, 2956, 2926, 2857, 1644, 1532, 1479, 1420, 1382, 1341, 1296, 1265, 1181, 1095, 988, 861, 789, 737 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₃H₁₇ClN₂O [M]⁺ 252.1029, found 252.1030.

***N*-Butyl-2-methylindoline-1-carboxamide (1l).** 1.46 g (75%); white solid; mp = 105–108 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.64 (d, *J* = 7.8 Hz, 1H), 7.16 (t, *J* = 7.8 Hz, 2H), 6.91 (t, *J* = 7.2 Hz, 1H), 4.74 (br s, 1H), 4.44–4.34 (m, 1H), 3.42–3.33 (m, 3H), 2.60 (d, *J* = 15.8 Hz, 1H), 1.61–1.51 (m, 2H), 1.45–1.33 (m, 2H), 1.30 (d, *J* = 6.3 Hz, 3H), 0.95 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 154.7, 142.4, 129.9, 127.5, 125.2, 121.9, 114.7, 55.2, 40.2, 36.3, 32.3, 21.0, 20.2, 13.8; IR (KBr) ν 3310, 2960, 2927, 2859, 1633, 1517, 1477, 1456, 1371, 1289, 1220, 1152, 1085, 1022, 933, 858, 753 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₄H₂₀N₂O [M]⁺ 232.1576, found 232.1574.

***N*-Butyl-3-methylindoline-1-carboxamide (1m).** 1.26 g (65%); white solid; mp = 89–91 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.85 (d, *J* = 8.0 Hz, 1H), 7.19–7.10 (m, 2H), 6.19 (td, *J* = 7.8 Hz, 1H), 4.58 (br s, 1H), 4.05 (t, *J* = 8.2 Hz, 1H), 3.51–3.39 (m, 2H), 3.32 (t, *J* = 7.0 Hz, 2H), 1.60–1.51 (m, 2H), 1.54–1.32 (m, 5H), 0.94 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 155.1, 143.2, 135.4, 127.7, 123.4, 121.7, 114.5, 55.2, 40.2, 34.6, 32.3, 20.3, 20.1, 13.8; IR (KBr) ν 3338, 2957, 2927, 2870, 1643, 1520, 1478, 1461, 1342, 1288, 1153, 1090, 1022, 930, 747 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₄H₂₀N₂O [M]⁺ 232.1576, found 232.1576.

***N*-Butylspiro[cyclohexane-1,3'-indoline]-1'-carboxamide (1n).** 1.68 g (70%); white solid; mp = 144–145 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.83 (d, *J* = 8.0 Hz, 1H), 7.19–7.08 (m, 2H), 6.92 (t, *J* = 8.2 Hz, 1H), 4.61 (br s, 1H), 3.69 (s, 2H), 3.34 (t, *J* = 7.1 Hz, 2H), 1.76–1.53 (m, 8H), 1.44–1.33 (m, 6H), 0.96 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 155.1, 142.6, 139.6, 127.9, 122.3, 121.7, 114.5, 57.5, 44.4, 40.3, 37.3, 32.3, 25.3, 23.0, 20.1, 13.8; IR (KBr) ν 3349, 2924, 2848, 1639, 1521, 1477, 1459, 1335, 1301, 1276, 1192, 1095, 1020, 936, 888, 750 cm⁻¹; HRMS (quadrupole, EI) calcd for C₁₈H₂₆N₂O [M]⁺ 286.2045, found 286.2039.

tert-Butyl 1-(Butylcarbamoyl)spiro[indoline-3,4'-piperidine]-1'-carboxylate (**1o**). 0.356 g (23%); light yellow sticky solid; ^1H NMR (300 MHz, CDCl_3) δ 7.78 (d, $J = 8.1$ Hz, 1H), 7.19 (td, $J = 8.6, 1.3$ Hz, 1H), 7.08 (d, $J = 7.4$ Hz, 1H), 6.94 (td, $J = 7.8, 0.9$ Hz, 1H), 4.68 (br s, 1H), 4.14–4.10 (m, 2H), 3.76 (s, 2H), 3.37–3.30 (m, 2H), 2.86 (t, $J = 12.8$ Hz, 2H), 1.88–1.78 (m, 2H), 1.66–1.36 (m, 15H), 0.96 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 155.0, 154.8, 142.7, 137.8, 128.4, 122.5, 122.0, 114.4, 79.8, 56.6, 42.7, 40.3, 36.5, 32.3, 29.7, 28.4, 20.1, 13.8; IR (KBr) ν 3325, 2925, 2847, 1638, 1520, 1477, 1456, 1341, 1279, 1220, 1152, 1095, 933, 754 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{22}\text{H}_{33}\text{N}_3\text{O}_3$ $[\text{M}]^+$ 387.2522, found 387.2521.

N-Butyl-3-methyl-3-phenylindoline-1-carboxamide (**1p**). 1.68 g (65%); light yellow solid; mp = 94–96 $^\circ\text{C}$; ^1H NMR (300 MHz, CDCl_3) δ 7.94 (d, $J = 8.1$ Hz, 1H), 7.32–7.19 (m, 6H), 6.94–6.93 (m, 2H), 4.51 (br s, 1H), 4.00 (d, $J = 9.0$ Hz, 1H), 3.88 (d, $J = 9.1$ Hz, 1H), 3.34–3.28 (m, 2H), 1.75 (s, 3H), 1.58–1.48 (m, 2H), 1.43–1.30 (m, 2H), 0.93 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 154.9, 147.0, 143.1, 138.6, 128.4, 128.1, 126.6, 126.4, 123.9, 122.1, 114.7, 64.1, 47.8, 40.2, 32.2, 27.2, 20.1, 13.8; IR (KBr) ν 3338, 2958, 2929, 2871, 1644, 1520, 1478, 1337, 1264, 1154, 1023, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{20}\text{H}_{24}\text{N}_2\text{O}$ $[\text{M}]^+$ 308.1889, found 308.1902.

N-Butyl-4a-methyl-2,3,4,4a-tetrahydro-1H-carbazole-9(9aH)-carboxamide (**1q**). 1.32 g (55%); white solid; mp = 164–166 $^\circ\text{C}$; ^1H NMR (300 MHz, CDCl_3) δ 7.61 (d, $J = 7.9$ Hz, 1H), 7.17 (td, $J = 8.1, 1.3$ Hz, 1H), 7.08 (d, $J = 7.2$ Hz, 1H), 6.95 (td, $J = 7.8, 0.8$ Hz, 1H), 4.77 (br s, 1H), 3.81–3.76 (m, 1H), 3.34 (s, 2H), 2.24–2.18 (m, 1H), 2.11–2.06 (m, 1H), 1.61–1.50 (m, 5H), 1.45–1.33 (m, 2H), 1.25–1.17 (m, 3H), 1.12 (s, 3H), 0.95 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 155.1, 141.6, 138.6, 127.5, 122.2, 121.8, 115.2, 67.3, 43.6, 40.2, 32.4, 32.3, 31.0, 28.3, 22.3, 21.8, 20.2, 13.8; IR (KBr) ν 3323, 2925, 2854, 1634, 1532, 1473, 1387, 1285, 1197, 1119, 1022, 928, 844, 749 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}$ $[\text{M}]^+$ 286.2045, found 286.2041.

N-Butyl-3,3-dimethyl-2-phenylindoline-1-carboxamide (**1r**). 1.35 g (50%); white solid; mp = 137–139 $^\circ\text{C}$; ^1H NMR (300 MHz, CDCl_3) δ 8.05 (d, $J = 8.0$ Hz, 1H), 7.33–7.28 (m, 3H), 7.24–7.20 (m, 1H), 7.16–7.13 (m, 2H), 7.06–6.95 (m, 2H), 4.71 (s, 1H), 4.30 (br s, 1H), 3.24–3.00 (m, 2H), 1.43 (s, 3H), 1.29–1.19 (m, 2H), 1.06–0.93 (m, 2H), 0.85 (s, 3H), 0.75 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 155.4, 143.1, 138.7, 138.2, 128.9, 128.3, 128.0, 126.6, 122.2, 122.1, 114.8, 45.4, 39.9, 32.6, 31.7, 24.2, 19.6, 13.6; IR (KBr) ν 3369, 2958, 2928, 2869, 1643, 1515, 1476, 1453, 1377, 1284, 1152, 1121, 1024, 939, 888, 826, 747 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{21}\text{H}_{26}\text{N}_2\text{O}$ $[\text{M}]^+$ 322.2045, found 322.2039.

Typical Procedure for the C7-Allylation of *N*-Butylindoline-1-carboxamides (3a–3t and 5b–5f). To an oven-dried sealed tube charged with *N*-butylindoline-1-carboxamide (**1f**) (65.5 mg, 0.3 mmol, 100 mol %), $[\text{RhCp}^*\text{Cl}_2]_2$ (4.6 mg, 2.5 mol %), AgSbF_6 (10.3 mg, 10 mol %), and $\text{Cu}(\text{OAc})_2$ (16.3 mg, 30 mol %) in *t*-AmOH (1 mL) was added allyl methyl carbonate (**2a**) (69.7 mg, 0.6 mmol, 200 mol %). The reaction mixture was allowed to stir at room temperature for 20 h. The reaction mixture was diluted with EtOAc (3 mL) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc = 7:1) to afford **3f** in 81% yield.

1-(7-Allylindolin-1-yl)ethanone (**3a**). 33.0 mg (55%); light yellow sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 7.07–7.02 (m, 3H), 5.86 (ddt, $J = 17.0, 10.1, 6.7$ Hz, 1H), 5.07–5.01 (m, 2H), 4.03 (s, 2H), 3.42 (d, $J = 6.3$ Hz, 2H), 3.00 (t, $J = 7.2$ Hz, 2H), 2.24 (s, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 168.6, 141.1, 136.8, 134.7, 129.0, 125.5, 125.0, 122.4, 116.0, 51.3, 38.4, 30.1, 23.9; IR (KBr) ν 2915, 1993, 1658, 1591, 1429, 1379, 1327, 1234, 1186, 1105, 968, 914, 847, 760, 730 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{13}\text{H}_{15}\text{NO}$ $[\text{M}]^+$ 201.1154, found 201.1151.

1-(7-Allylindolin-1-yl)-2,2-dimethylpropan-1-one (**3b**). 26.7 mg (37%); light yellow sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 7.08–7.07 (m, 1H), 7.04–7.01 (m, 2H), 5.82 (ddt, $J = 17.0, 10.0, 6.7$ Hz, 1H), 5.08–5.01 (m, 2H), 4.08 (t, $J = 7.3$ Hz, 2H), 3.28 (d, $J = 6.7$ Hz, 2H), 3.00 (t, $J = 7.3$ Hz, 2H), 1.36 (s, 9H); ^{13}C NMR (175 MHz,

CDCl_3) δ 178.2, 143.0, 136.7, 134.5, 131.6, 128.5, 125.2, 122.3, 116.1, 51.3, 40.1, 38.3, 31.3, 28.7; IR (KBr) ν 2963, 1967, 1648, 1587, 1430, 1347, 1322, 1265, 1183, 1149, 1088, 966, 903, 730 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{16}\text{H}_{21}\text{NO}$ $[\text{M}]^+$ 243.1623, found 243.1622.

7-Allyl-*N,N*-dimethylindoline-1-carboxamide (**3c**). 24.2 mg (35%); light brown sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 7.02 (d, $J = 7.2$ Hz, 1H), 6.97 (d, $J = 7.1$ Hz, 1H), 6.93 (t, $J = 7.4$ Hz, 1H), 5.82 (ddt, $J = 17.0, 10.0, 6.8$ Hz, 1H), 5.08–5.00 (m, 2H), 3.88–3.84 (m, 2H), 3.27 (d, $J = 6.8$ Hz, 2H), 3.05 (t, $J = 7.7$ Hz, 2H), 2.95 (s, 6H); ^{13}C NMR (175 MHz, CDCl_3) δ 161.3, 143.6, 136.5, 133.6, 131.5, 128.3, 123.7, 122.4, 116.0, 52.9, 50.5, 38.3, 37.4, 30.6; IR (KBr) ν 2924, 2010, 1650, 1482, 1451, 1377, 1292, 1258, 1166, 1062, 1006, 911, 731 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{14}\text{H}_{18}\text{N}_2\text{O}$ $[\text{M}]^+$ 230.1419, found 230.1424.

(7-Allylindolin-1-yl)(pyrrolidin-1-yl)methanone (**3d**). 9.1 mg (12%); light yellow sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 6.98 (d, $J = 6.9$ Hz, 1H), 6.92 (d, $J = 7.5$ Hz, 1H), 6.87 (t, $J = 7.4$ Hz, 1H), 5.75 (ddt, $J = 16.9, 10.0, 6.8$ Hz, 1H), 5.02–4.92 (m, 2H), 3.87 (t, $J = 8.2$ Hz, 1H), 3.82 (t, $J = 8.0$ Hz, 1H), 3.39–3.36 (m, 4H), 3.27 (d, $J = 6.7$ Hz, 2H), 3.02–2.97 (m, 2H), 1.83–1.81 (m, 4H); ^{13}C NMR (175 MHz, CDCl_3) δ 159.3, 143.5, 136.9, 131.4, 128.6, 123.6, 122.5, 115.8, 52.2, 50.2, 48.0, 37.8, 30.6, 28.7, 25.7; IR (KBr) ν 2874, 1966, 1644, 1592, 1452, 1383, 1285, 1254, 1196, 1124, 1048, 994, 912, 853, 753 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}$ $[\text{M}]^+$ 256.1576, found 256.1573.

7-Allyl-*N*-phenylindoline-1-carboxamide (**3e**). 15.9 mg (19%); light yellow solid; mp = 178–179 $^\circ\text{C}$; ^1H NMR (700 MHz, CDCl_3) δ 7.35 (d, $J = 8.6, 1.1$ Hz, 2H), 7.28 (t, $J = 7.4$ Hz, 2H), 7.13–7.02 (m, 4H), 6.54 (s, 1H), 5.87 (ddt, $J = 16.9, 10.2, 6.5$ Hz, 1H), 5.08–5.04 (m, 2H), 4.16 (t, $J = 7.2$ Hz, 2H), 3.41 (d, $J = 6.5$ Hz, 2H), 3.03 (t, $J = 7.6$ Hz, 2H); ^{13}C NMR (175 MHz, CDCl_3) δ 154.3, 141.5, 138.5, 136.3, 135.6, 129.3, 129.2, 129.0, 125.2, 123.5, 123.3, 119.6, 116.8, 52.0, 36.9, 30.2; IR (KBr) ν 3263, 3051, 2926, 1998, 1650, 1594, 1525, 1479, 1439, 1343, 1315, 1240, 1141, 1025, 916, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{18}\text{H}_{18}\text{N}_2\text{O}$ $[\text{M}]^+$ 278.1419, found 278.1422.

7-Allyl-*N*-butylindoline-1-carboxamide (**3f**). 65.0 mg (81%); light brown solid; mp = 77–81 $^\circ\text{C}$; ^1H NMR (700 MHz, CDCl_3) δ 7.06 (d, $J = 7.1$ Hz, 1H), 7.02 (d, $J = 7.6$ Hz, 1H), 6.98 (t, $J = 7.4$ Hz, 1H), 5.90 (ddt, $J = 17.0, 10.1, 6.4$ Hz, 1H), 5.10–5.06 (m, 2H), 4.70 (br s, 1H), 4.06 (t, $J = 7.6$ Hz, 2H), 3.37 (t, $J = 6.4$ Hz, 2H), 3.25–3.22 (m, 2H), 2.96 (t, $J = 7.7$ Hz, 2H), 1.49–1.45 (m, 2H), 1.34–1.29 (m, 2H), 0.89 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.3, 142.2, 136.8, 135.5, 129.1, 128.7, 124.6, 123.0, 116.3, 51.9, 40.7, 36.9, 32.4, 30.1, 20.3, 14.0; IR (KBr) ν 3313, 3051, 2929, 1993, 1642, 1522, 1482, 1450, 1311, 1248, 1187, 1152, 1086, 912, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{16}\text{H}_{22}\text{N}_2\text{O}$ $[\text{M}]^+$ 258.1732, found 258.1735.

7-Allyl-*N*-butyl-4-methylindoline-1-carboxamide (**3g**). 41.3 mg (44%); brown solid; mp = 89–92 $^\circ\text{C}$; ^1H NMR (700 MHz, CDCl_3) δ 6.89 (s, 1H), 6.82 (s, 1H), 5.91 (ddt, $J = 17.0, 10.1, 6.4$ Hz, 1H), 5.10–5.06 (m, 2H), 4.71 (br s, 1H), 4.05 (t, $J = 7.6$ Hz, 2H), 3.33 (d, $J = 6.3$ Hz, 2H), 3.23–3.20 (m, 2H), 2.90 (t, $J = 7.6$ Hz, 2H), 2.64 (s, 3H), 1.47–1.43 (m, 2H), 1.33–1.28 (m, 2H), 0.89 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.5, 139.9, 136.8, 135.8, 134.3, 129.4, 128.3, 123.9, 116.3, 52.0, 40.7, 36.7, 32.4, 30.1, 21.1, 20.2, 13.9; IR (KBr) ν 3314, 3076, 2927, 1975, 1639, 1527, 1474, 1316, 1244, 1191, 1124, 1080, 994, 909, 854, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{N}_2\text{O}$ $[\text{M}]^+$ 272.1889, found 272.1885.

7-Allyl-5-bromo-*N*-butylindoline-1-carboxamide (**3h**). 77.0 mg (78%); brown solid; mp = 126–130 $^\circ\text{C}$; ^1H NMR (700 MHz, CDCl_3) δ 7.15 (s, 1H), 7.12 (s, 1H), 5.84 (ddt, $J = 16.8, 10.2, 6.5$ Hz, 1H), 5.10–5.07 (m, 2H), 4.68 (br s, 1H), 4.01 (t, $J = 7.7$ Hz, 2H), 3.31 (d, $J = 6.5$ Hz, 2H), 3.23–3.20 (m, 2H), 2.95 (t, $J = 7.7$ Hz, 2H), 1.48–1.44 (m, 2H), 1.34–1.28 (m, 2H), 0.89 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 156.8, 141.4, 136.9, 135.8, 131.4, 130.7, 125.7, 116.9, 116.8, 51.7, 40.6, 36.7, 32.2, 29.7, 20.0, 13.8; IR (KBr) ν 3308, 3066, 2929, 1995, 1640, 1530, 1454, 1422, 1307, 1244, 1187,

1091, 993, 912, 853, 736 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{16}\text{H}_{21}\text{BrN}_2\text{O}$ $[\text{M}]^+$ 336.0837, found 336.0833.

7-Allyl-N-butyl-5-chloroindoline-1-carboxamide (3i). 69.4 mg (79%); light yellow solid; mp = 112–117 °C; ^1H NMR (700 MHz, CDCl_3) δ 7.02 (s, 1H), 6.99 (s, 1H), 5.85 (ddt, J = 17.5, 10.3, 6.5 Hz, 1H), 5.11–5.08 (m, 2H), 4.62 (t, J = 5.1 Hz, 1H), 4.04 (t, J = 7.7 Hz, 2H), 3.32 (d, J = 6.5 Hz, 2H), 3.25–3.22 (m, 2H), 2.96 (t, J = 7.7 Hz, 2H), 1.49–1.45 (m, 2H), 1.35–1.29 (m, 2H), 0.90 (t, J = 7.3 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.1, 141.0, 136.8, 136.0, 130.4, 129.5, 128.7, 123.0, 117.0, 52.0, 40.8, 36.9, 32.4, 30.0, 20.2, 13.9; IR (KBr) ν 3297, 3065, 2928, 1991, 1639, 1531, 1456, 1424, 1308, 1254, 1188, 1145, 1092, 992, 913, 857, 736 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{16}\text{H}_{21}\text{ClN}_2\text{O}$ $[\text{M}]^+$ 292.1342, found 292.1335.

7-Allyl-N-butyl-6-fluoroindoline-1-carboxamide (3j). 68.7 mg (83%); light yellow solid; mp = 76–81 °C; ^1H NMR (700 MHz, CDCl_3) δ 6.97 (t, J = 7.2 Hz, 1H), 6.68 (t, J = 9.9 Hz, 1H), 5.90 (ddt, J = 16.8, 10.2, 5.8 Hz, 1H), 4.99–4.96 (m, 2H), 4.83 (br s, 1H), 4.05 (t, J = 7.7 Hz, 2H), 3.41 (d, J = 5.8 Hz, 2H), 3.21–3.18 (m, 2H), 2.89 (t, J = 7.5 Hz, 2H), 1.47–1.42 (m, 2H), 1.32–1.26 (m, 2H), 0.87 (t, J = 7.2 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 160.6 (d, $J_{\text{C-F}}$ = 240.9 Hz), 156.8, 143.6 (d, $J_{\text{C-F}}$ = 7.6 Hz), 135.3, 130.4, 123.0 (d, $J_{\text{C-F}}$ = 10.2 Hz), 116.7 (d, $J_{\text{C-F}}$ = 20.1 Hz), 115.2, 110.5 (d, $J_{\text{C-F}}$ = 23.9 Hz), 52.5, 40.5, 32.1, 30.5, 30.4, 29.3, 20.0, 13.7; IR (KBr) ν 3310, 3071, 2930, 1967, 1644, 1525, 1470, 1309, 1240, 1215, 1139, 1113, 992, 910, 802, 735 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{16}\text{H}_{21}\text{FN}_2\text{O}$ $[\text{M}]^+$ 276.1638, found 276.1636.

7-Allyl-N-butyl-6-chloroindoline-1-carboxamide (3k). 77.0 mg (82%); light yellow solid; mp = 87–93 °C; ^1H NMR (700 MHz, CDCl_3) δ 7.05 (d, J = 7.9 Hz, 1H), 6.99 (d, J = 7.9 Hz, 1H), 5.87 (ddt, J = 17.1, 10.1, 5.8 Hz, 1H), 5.01–4.99 (m, 1H), 4.96–4.93 (m, 1H), 4.78 (br s, 1H), 4.06 (t, J = 7.7 Hz, 2H), 3.55 (d, J = 7.4 Hz, 2H), 3.22–3.19 (m, 2H), 2.91 (t, J = 7.7 Hz, 2H), 1.47–1.43 (m, 2H), 1.32–1.27 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.1, 144.1, 134.8, 134.4, 134.0, 127.1, 125.7, 123.5, 115.5, 52.6, 40.6, 34.2, 32.1, 29.7, 20.0, 13.8; IR (KBr) ν 3313, 3055, 2956, 1963, 1647, 1527, 1445, 1427, 1331, 1303, 1265, 1190, 1087, 990, 912, 801, 733 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{16}\text{H}_{21}\text{ClN}_2\text{O}$ $[\text{M}]^+$ 292.1342, found 292.1344.

7-Allyl-N-butyl-2-methylindoline-1-carboxamide (3l). 68.1 mg (83%); light yellow sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 7.05 (d, J = 7.0 Hz, 1H), 7.00 (d, J = 7.4 Hz, 1H), 6.97 (t, J = 7.3 Hz, 1H), 5.91–5.85 (m, 1H), 5.06–5.03 (m, 2H), 4.70 (br s, 1H), 4.64–4.60 (m, 1H), 3.43–3.40 (m, 1H), 3.36–3.28 (m, 2H), 3.27–3.17 (m, 2H), 2.36 (d, J = 6.6 Hz, 1H), 1.48–1.43 (m, 2H), 1.32–1.28 (m, 2H), 1.17 (d, J = 6.7 Hz, 3H), 0.88 (t, J = 7.2 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 156.7, 140.5, 136.6, 134.2, 129.1, 128.8, 124.4, 123.4, 115.9, 58.6, 40.4, 36.9, 36.7, 32.2, 21.7, 20.1, 13.8; IR (KBr) ν 3348, 2957, 1969, 1641, 1513, 1445, 1377, 1357, 1298, 1264, 1190, 1108, 1075, 994, 910, 735 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{N}_2\text{O}$ $[\text{M}]^+$ 272.1889, found 272.1885.

7-Allyl-N-butyl-3-methylindoline-1-carboxamide (3m). 62.6 mg (77%); light yellow solid; mp = 72–76 °C; ^1H NMR (700 MHz, CDCl_3) δ 7.02–6.98 (m, 3H), 5.90 (ddt, J = 16.8, 10.1, 6.3 Hz, 1H), 5.09–5.05 (m, 2H), 4.81 (br s, 1H), 4.22 (t, J = 8.8 Hz, 1H), 3.55–3.52 (m, 1H), 3.36 (d, J = 6.3 Hz, 2H), 3.30–3.17 (m, 3H), 1.47–1.43 (m, 2H), 1.32–1.27 (m, 2H), 1.20 (d, J = 6.8 Hz, 3H), 0.88 (t, J = 7.2 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.3, 141.7, 140.4, 136.6, 128.9, 128.5, 124.6, 121.5, 116.1, 59.5, 40.5, 36.6, 36.5, 32.2, 20.0, 18.4, 13.8; IR (KBr) ν 3312, 2958, 1959, 1641, 1524, 1443, 1321, 1300, 1243, 1144, 1089, 1059, 993, 910, 735 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{N}_2\text{O}$ $[\text{M}]^+$ 272.1889, found 272.1886.

7'-Allyl-N-butylspiro[cyclohexane-1,3'-indoline]-1'-carboxamide (3n). 77.0 mg (79%); light yellow solid; mp = 140–146 °C; ^1H NMR (700 MHz, CDCl_3) δ 7.03–6.98 (m, 3H), 5.90 (ddt, J = 17.0, 10.1, 6.4 Hz, 1H), 5.11–5.06 (m, 2H), 4.79 (br s, 1H), 3.89 (s, 2H), 3.36 (d, J = 6.4 Hz, 2H), 3.24–3.21 (m, 2H), 1.71–1.67 (m, 4H), 1.56–1.50 (m, 4H), 1.47–1.43 (m, 2H), 1.41–1.35 (m, 2H), 1.32–1.26 (m, 2H), 0.88 (t, J = 7.3 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.6, 144.4, 141.3, 136.5, 129.0, 128.5, 124.7, 120.5, 116.1, 61.3, 46.0, 40.5, 36.4, 35.6, 32.2, 25.6, 23.2, 20.2, 13.8; IR (KBr) ν 3349, 2927, 1994, 1642,

1518, 1442, 1311, 1242, 1146, 1065, 994, 907, 735 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{21}\text{H}_{30}\text{N}_2\text{O}$ $[\text{M}]^+$ 326.2358, found 326.2359.

tert-Butyl 7-Allyl-1-(butylcarbamoyl)spiro[indoline-3,4'-piperidine]-1'-carboxylate (3o). 79.9 mg (62%); light brown sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 7.08–7.04 (m, 2H), 7.00–6.97 (m, 1H), 5.92 (ddt, J = 16.6, 10.2, 6.3 Hz, 1H), 5.11–5.09 (m, 2H), 4.77 (br s, 1H), 4.14–3.97 (m, 5H), 3.36 (d, J = 6.3 Hz, 2H), 3.24–3.21 (m, 2H), 2.88 (br s, 2H), 1.74 (br s, 2H), 1.54–1.46 (m, 3H), 1.45 (s, 9H), 1.32–1.25 (m, 2H), 0.88 (t, J = 7.3 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.4, 154.8, 142.7, 141.3, 136.3, 129.6, 128.7, 125.1, 120.7, 116.4, 79.7, 60.2, 44.4, 40.5, 36.2, 32.2, 28.4, 25.1, 20.0, 13.7; IR (KBr) ν 3312, 2929, 1967, 1668, 1525, 1424, 1365, 1237, 1169, 1144, 1091, 969, 911, 862, 732 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{25}\text{H}_{37}\text{N}_3\text{O}_3$ $[\text{M}]^+$ 427.2835, found 427.2833.

7-Allyl-N-butyl-3-methyl-3-phenylindoline-1-carboxamide (3p). 81.8 mg (78%); light brown solid; mp = 79–83 °C; ^1H NMR (700 MHz, CDCl_3) δ 7.27–7.22 (m, 4H), 7.18 (t, J = 8.4 Hz, 1H), 7.12 (d, J = 7.6 Hz, 1H), 7.07 (t, J = 7.4 Hz, 1H), 6.92 (d, J = 7.2 Hz, 1H), 5.98 (ddt, J = 17.0, 10.1, 6.4 Hz, 1H), 5.16–5.11 (m, 2H), 4.81 (br s, 1H), 4.30 (d, J = 10.8 Hz, 1H), 3.99 (d, J = 10.7 Hz, 1H), 3.43 (d, J = 6.5 Hz, 2H), 3.21–3.12 (m, 2H), 1.65 (s, 3H), 1.37–1.32 (m, 2H), 1.22–1.17 (m, 2H), 0.84 (t, J = 7.3 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.3, 145.8, 143.4, 141.9, 136.5, 129.3, 129.0, 128.3, 126.6, 126.5, 125.1, 122.2, 116.3, 67.2, 49.8, 40.3, 36.5, 32.2, 25.5, 19.9, 13.8; IR (KBr) ν 3312, 2958, 2929, 2010, 1644, 1524, 1441, 1317, 1263, 1068, 1029, 912, 758, 735 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{23}\text{H}_{28}\text{N}_2\text{O}$ $[\text{M}]^+$ 348.2202, found 348.2204.

8-Allyl-N-butyl-4a-methyl-2,3,4,4a-tetrahydro-1H-carbazole-9-(9aH)-carboxamide (3q). 76.1 mg (78%); light yellow solid; mp = 94–101 °C; ^1H NMR (700 MHz, CDCl_3) δ 7.05–7.02 (m, 2H), 6.94 (dd, J = 6.8, 1.6 Hz, 1H), 5.89 (ddt, J = 17.0, 10.2, 6.8 Hz, 1H), 5.08–5.05 (m, 2H), 4.70 (t, J = 5.3 Hz, 1H), 4.13–4.10 (m, 1H), 3.44–3.28 (m, 2H), 3.25–3.23 (m, 2H), 2.19–2.16 (m, 1H), 2.03–1.99 (m, 1H), 1.53–1.23 (m, 10H), 1.08 (s, 3H), 0.88 (t, J = 7.3 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.5, 142.9, 140.9, 136.7, 130.0, 129.2, 125.2, 120.3, 116.1, 70.7, 45.6, 40.6, 36.4, 33.3, 32.5, 30.6, 30.0, 23.6, 22.2, 20.2, 13.9; IR (KBr) ν 3288, 2926, 1994, 1638, 1524, 1436, 1304, 1264, 1187, 1146, 1083, 993, 911, 757, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{21}\text{H}_{30}\text{N}_2\text{O}$ $[\text{M}]^+$ 326.2358, found 326.2359.

7-Allyl-N-butyl-3,3-dimethyl-2-phenylindoline-1-carboxamide (3r). 87.3 mg (80%); yellow sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 7.26–7.23 (m, 3H), 7.11 (d, J = 7.7 Hz, 1H), 7.06–7.03 (m, 3H), 6.91 (dd, J = 7.2, 1.1 Hz, 1H), 5.98 (ddt, J = 16.9, 10.0, 6.7 Hz, 1H), 5.17–5.08 (m, 2H), 4.94 (s, 1H), 4.54 (t, J = 5.3 Hz, 1H), 3.61 (dd, J = 15.8, 6.6 Hz, 1H), 3.46 (dd, J = 15.8, 6.7 Hz, 1H), 3.30–3.26 (m, 1H), 3.21–3.16 (m, 1H), 1.48–1.43 (m, 2H), 1.42 (s, 3H), 1.31–1.25 (m, 2H), 0.89 (t, J = 7.3 Hz, 6H); ^{13}C NMR (175 MHz, CDCl_3) δ 157.0, 142.0, 141.2, 140.0, 136.8, 129.4, 128.4, 128.1, 127.7, 126.8, 124.5, 120.5, 116.2, 78.4, 46.3, 40.3, 37.4, 32.5, 32.1, 22.9, 19.9, 13.7; IR (KBr) ν 3346, 2957, 2928, 2007, 1945, 1659, 1505, 1439, 1362, 1290, 1246, 1203, 1138, 1072, 994, 910, 751 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{24}\text{H}_{30}\text{N}_2\text{O}$ $[\text{M}]^+$ 362.2358, found 362.2368.

(E)-7-(But-2-enyl)-N-butylindoline-1-carboxamide and (Z)-7-(But-2-enyl)-N-butylindoline-1-carboxamide (5b). 72.6 mg (89%); brown solid; mp = 85–89 °C; $E:Z$ ratio = 1:2.7; ^1H NMR (700 MHz, CDCl_3) E -isomer, δ 7.05 (d, J = 7.0 Hz, 1H), 7.02 (t, J = 7.1 Hz, 1H), 6.99–6.96 (m, 1H), 5.60–5.45 (m, 2H), 4.70 (br s, 1H), 4.08–4.05 (m, 2H), 3.54 (d, J = 7.0 Hz, 2H), 3.26–3.21 (m, 2H), 2.96–2.93 (m, 2H), 1.67–1.66 (m, 3H), 1.50–1.43 (m, 2H), 1.35–1.28 (m, 2H), 0.90 (t, J = 7.3 Hz, 3H); Z -isomer, δ 7.05 (d, J = 7.0 Hz, 1H), 7.02 (t, J = 7.1 Hz, 1H), 6.99–6.96 (m, 1H), 5.60–5.45 (m, 2H), 4.74 (br s, 1H), 4.08–4.05 (m, 2H), 3.28 (br s, 2H), 3.26–3.21 (m, 2H), 2.96–2.93 (m, 2H), 1.67–1.66 (m, 3H), 1.50–1.43 (m, 2H), 1.35–1.28 (m, 2H), 0.89 (t, J = 7.3 Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) E -isomer, δ 157.4, 142.2, 135.4, 129.5, 129.2, 128.4, 125.8, 124.8, 122.7, 51.9, 40.7, 35.5, 32.4, 30.2, 20.3, 18.1, 13.9; Z -isomer, δ 157.3, 142.1, 135.6, 130.2, 129.1, 128.6, 126.9, 124.6, 122.8, 51.9, 40.7, 35.5, 32.4, 30.1, 20.3, 18.1, 13.9; IR (KBr) ν 3312, 2956, 2928, 2011, 1641, 1525, 1449,

1308, 1248, 1188, 1152, 1089, 966, 847, 757 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{N}_2\text{O}$ $[\text{M}]^+$ 272.1889, found 272.1884.

(*E*)-*N*-Butyl-7-(undec-2-enyl)indoline-1-carboxamide (**5c**). 93.7 mg (84%); orange sticky solid; *E*:*Z* ratio = 1:2.9; ^1H NMR (700 MHz, CDCl_3) *E*-isomer, δ 7.06–7.01 (m, 2H), 6.98–6.96 (m, 1H), 5.53–5.41 (m, 2H), 4.70 (br s, 1H), 4.07 (t, $J = 7.6$ Hz, 2H), 3.34 (d, $J = 7.0$ Hz, 2H), 3.26–3.21 (m, 2H), 2.96–2.93 (m, 2H), 2.10–2.07 (m, 2H), 1.50–1.44 (m, 2H), 1.40–1.06 (m, 14H), 0.91 (t, $J = 7.3$ Hz, 3H), 0.85 (t, $J = 7.0$ Hz, 3H); *Z*-isomer, δ 7.06–7.01 (m, 2H), 6.98–6.96 (m, 1H), 5.53–5.41 (m, 2H), 4.74 (br s, 1H), 4.07 (t, $J = 7.6$ Hz, 2H), 3.29 (d, $J = 5.3$ Hz, 2H), 3.26–3.21 (m, 2H), 2.96–2.93 (m, 2H), 2.00–1.98 (m, 2H), 1.50–1.44 (m, 2H), 1.40–1.06 (m, 14H), 0.89 (t, $J = 7.3$ Hz, 3H), 0.85 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) *E*-isomer, δ 157.4, 142.1, 135.3, 132.0, 130.3, 128.6, 127.3, 124.7, 122.7, 51.9, 40.7, 35.5, 32.4, 32.0, 30.2, 29.8, 29.7, 29.5, 29.4, 27.5, 22.8, 20.3, 13.9; *Z*-isomer, δ 157.3, 142.1, 135.5, 132.7, 129.6, 129.0, 127.8, 124.6, 122.8, 51.9, 40.7, 35.5, 32.7, 32.0, 30.1, 29.8, 29.6, 29.5, 29.4, 27.5, 22.8, 20.3, 14.2; IR (KBr) ν 3313, 2929, 2853, 1966, 1644, 1527, 1450, 1313, 1249, 1188, 969, 850, 763, 735 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{24}\text{H}_{38}\text{N}_2\text{O}$ $[\text{M}]^+$ 370.2984, found 370.2980.

(*S,E*)-*N*-Butyl-7-(5,9-dimethyldeca-2,8-dienyl)indoline-1-carboxamide and (*S,Z*)-*N*-Butyl-7-(5,9-dimethyldeca-2,8-dienyl)indoline-1-carboxamide (**5d**). 96.7 mg (84%); orange sticky solid; *E*:*Z* ratio = 1:2.8; ^1H NMR (700 MHz, CDCl_3) *E*-isomer, δ 7.06–7.02 (m, 2H), 6.97 (t, $J = 7.4$ Hz, 1H), 5.53–5.44 (m, 2H), 5.08–5.05 (m, 1H), 4.67 (t, $J = 5.6$ Hz, 1H), 4.08–4.05 (m, 2H), 3.34 (d, $J = 5.8$ Hz, 2H), 3.26–3.22 (m, 2H), 2.95 (t, $J = 7.5$ Hz, 2H), 2.11–1.82 (m, 4H), 1.66 (s, 3H), 1.57 (s, 3H), 1.50–1.44 (m, 3H), 1.37–1.29 (m, 3H), 1.16–1.08 (m, 1H), 0.91–0.85 (m, 6H); *Z*-isomer, δ 7.06–7.02 (m, 2H), 6.97 (t, $J = 7.4$ Hz, 1H), 5.53–5.44 (m, 2H), 5.08–5.05 (m, 1H), 4.70 (t, $J = 5.5$ Hz, 1H), 4.08–4.05 (m, 2H), 3.31 (d, $J = 4.6$ Hz, 2H), 3.26–3.22 (m, 2H), 2.95 (t, $J = 7.5$ Hz, 2H), 2.11–1.82 (m, 4H), 1.66 (s, 3H), 1.57 (s, 3H), 1.50–1.44 (m, 3H), 1.37–1.29 (m, 3H), 1.16–1.08 (m, 1H), 0.91–0.85 (m, 6H); ^{13}C NMR (175 MHz, CDCl_3) *E*-isomer, δ 157.3, 142.1, 135.5, 131.3, 131.2, 130.5, 129.8, 129.1, 129.0, 128.6, 128.1, 125.1, 125.0, 124.8, 124.6, 122.8, 122.7, 51.9, 40.8, 36.9, 34.8, 33.2, 32.5, 30.4, 25.8, 20.3, 19.7, 17.8, 14.0; *Z*-isomer, δ 157.3, 142.1, 135.5, 131.2, 130.5, 129.8, 129.1, 128.6, 125.0, 124.6, 122.8, 51.9, 40.1, 36.8, 35.7, 34.8, 32.9, 32.5, 30.4, 30.1, 25.9, 25.7, 20.3, 19.6, 17.8, 14.0; IR (KBr) ν 3315, 2925, 1975, 1643, 1527, 1449, 1312, 1249, 1188, 1089, 970, 847, 762, 737 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{25}\text{H}_{38}\text{N}_2\text{O}$ $[\text{M}]^+$ 382.2984, found 382.2982.

N-Butyl-7-cinnamylindoline-1-carboxamide (**5e**). 85.3 mg (85%); light yellow solid; mp = 121–123 $^{\circ}\text{C}$; *E*:*Z* ratio = 50>:1; ^1H NMR (700 MHz, CDCl_3) *E*-isomer, δ 7.32 (t, $J = 8.4$ Hz, 2H), 7.26 (t, $J = 7.6$ Hz, 2H), 7.19–7.16 (m, 1H), 7.09–7.08 (m, 2H), 6.99 (t, $J = 7.5$ Hz, 1H), 6.45 (d, $J = 15.8$ Hz, 1H), 6.28 (dt, $J = 15.8, 6.7$ Hz, 1H), 4.67 (br s, 1H), 4.07 (t, $J = 7.8$ Hz, 2H), 3.55 (dd, $J = 6.7, 1.2$ Hz, 2H), 3.25–3.22 (m, 2H), 2.99 (t, $J = 7.7$ Hz, 2H), 1.47–1.42 (m, 2H), 1.32–1.26 (m, 2H), 0.86 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) *E*-isomer, δ 157.4, 142.3, 137.7, 135.4, 131.6, 129.3, 129.1, 128.7, 128.6, 127.3, 126.3, 124.7, 123.1, 51.9, 40.9, 36.3, 32.5, 30.2, 20.3, 14.0; IR (KBr) ν 3312, 2926, 1996, 1641, 1525, 1449, 1311, 1247, 1188, 963, 752, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{22}\text{H}_{26}\text{N}_2\text{O}$ $[\text{M}]^+$ 334.2045, found 334.2039.

Experimental Procedure and Characterization for the C7-Allylation of Indoline with 2-Vinyloxirane. To an oven-dried sealed tube charged with *N*-butylindoline-1-carboxamide (**1f**) (65.5 mg, 0.3 mmol, 100 mol %), $[\text{RhCp}^*\text{Cl}_2]_2$ (4.6 mg, 2.5 mol %), AgSbF_6 (10.3 mg, 10 mol %), and $\text{Cu}(\text{OAc})_2$ (16.3 mg, 30 mol %) in *t*-AmOH (1 mL) was added 2-vinyloxirane (**2g**) (42.1 mg, 0.6 mmol, 200 mol %). The reaction mixture was allowed to stir at room temperature for 20 h. The reaction mixture was diluted with EtOAc (3 mL) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc = 1:1) to afford **5g** in 69% yield.

N-Butyl-7-(4-hydroxybut-2-enyl)indoline-1-carboxamide (**5g**). Brown sticky solid; *E*:*Z* ratio = 3:1; ^1H NMR (700 MHz, CDCl_3)

E-isomer, δ 7.07–7.00 (m, 2H), 6.97 (t, $J = 7.3$ Hz, 1H), 5.77–5.67 (m, 2H), 4.66 (br s, 1H), 4.08 (d, $J = 5.6$ Hz, 2H), 4.04 (t, $J = 7.7$ Hz, 2H), 3.38 (d, $J = 6.1$ Hz, 2H), 3.27–3.23 (m, 2H), 2.97 (t, $J = 7.7$ Hz, 2H), 1.65 (br s, 1H), 1.52–1.45 (m, 2H), 1.37–1.29 (m, 2H), 0.90 (t, $J = 7.3$ Hz, 3H); *Z*-isomer, δ 7.07–7.00 (m, 2H), 6.97 (t, $J = 7.3$ Hz, 1H), 5.62–5.58 (m, 2H), 4.62 (br s, 1H), 4.17 (d, $J = 7.0$ Hz, 2H), 4.01 (t, $J = 7.7$ Hz, 2H), 3.43 (d, $J = 7.1$ Hz, 2H), 3.27–3.23 (m, 2H), 3.00 (t, $J = 7.7$ Hz, 2H), 1.65 (br s, 1H), 1.52–1.45 (m, 2H), 1.37–1.29 (m, 2H), 0.91 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) *E*-isomer, δ 157.4, 142.1, 135.4, 130.7, 129.6, 129.1, 128.8, 124.6, 123.1, 63.7, 58.4, 51.9, 40.8, 35.7, 32.4, 30.1, 20.3, 14.0; *Z*-isomer, δ 157.6, 141.9, 134.8, 130.9, 129.6, 129.0, 128.8, 124.7, 122.8, 63.7, 58.4, 51.8, 40.8, 35.7, 32.4, 30.1, 20.3, 14.0; IR (KBr) ν 3313, 2929, 2870, 1946, 1642, 1523, 1450, 1314, 1249, 1187, 1091, 972, 905, 762, 731 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{N}_2\text{O}_2$ $[\text{M}]^+$ 288.1838, found 288.1837.

Experimental Procedure and Characterization for Olefin Metathesis of 3f. To an oven-dried sealed tube charged with 7-allyl-*N*-butylindoline-1-carboxamide (**3f**) (51.6 mg, 0.2 mmol, 100 mol %) in toluene (1 mL) were added Hoveyda–Grubbs second generation catalyst (8.4 mg, 5 mol %) and ethyl acrylate (60.0 mg, 0.6 mmol, 300 mol %) at room temperature. The reaction mixture was allowed to stir at 100 $^{\circ}\text{C}$ for 24 h, and cooled to room temperature. The reaction mixture was diluted with EtOAc (3 mL) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc = 5:1) to afford 40.0 mg of **6a** in 60% yield.

(*E*)-Ethyl 4-(1-(butylcarbamoyl)indolin-7-yl)but-2-enoate (**6a**). Light brown sticky solid; *E*:*Z* ratio = 50>:1; ^1H NMR (700 MHz, CDCl_3) δ 7.07 (d, $J = 6.7$ Hz, 1H), 7.01–6.94 (m, 3H), 5.81 (d, $J = 15.5$ Hz, 1H), 4.59 (t, $J = 5.3$ Hz, 1H), 4.15–4.12 (m, 2H), 4.02 (t, $J = 7.8$ Hz, 2H), 3.56 (d, $J = 6.7$ Hz, 2H), 3.25–3.22 (m, 2H), 3.00 (t, $J = 7.7$ Hz, 2H), 1.49–1.45 (m, 2H), 1.35–1.30 (m, 2H), 1.25–1.22 (m, 3H), 0.90 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 166.7, 157.1, 147.1, 142.3, 135.0, 129.2, 127.1, 124.6, 123.3, 122.5, 60.3, 51.7, 40.8, 35.9, 32.4, 30.1, 29.8, 20.2, 14.4, 13.9; IR (KBr) ν 3328, 2927, 2975, 1715, 1646, 1523, 1459, 1306, 1263, 1187, 1154, 1042, 983, 905, 845, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{19}\text{H}_{26}\text{N}_2\text{O}_3$ $[\text{M}]^+$ 330.1943, found 330.1943.

Experimental Procedure and Characterization for the Intramolecular Cyclization of 6a. To a stirred solution of (*E*)-ethyl 4-(1-(butylcarbamoyl)indolin-7-yl)but-2-enoate (**6a**) (44.4 mg, 0.13 mmol, 100 mol %) in DMF (2 mL) was added NaH (7.8 mg, 0.19 mmol, 150 mol %, 60% dispersion in mineral oil) at 0 $^{\circ}\text{C}$. The reaction mixture was stirred for 1 h at 0 $^{\circ}\text{C}$. The reaction mixture was quenched and partitioned between EtOAc and H_2O . The organic layer was dried over MgSO_4 and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc = 6:1) to afford 18.6 mg of **6b** in 45% yield.

Ethyl 3-butyl-4-oxo-1,2,3,4,6,7-hexahydro-[1,3]diazepino[6,7,1-h]indole-2-carboxylate (**6b**). Light brown sticky solid; ^1H NMR (700 MHz, CDCl_3) δ 7.02 (d, $J = 7.0$ Hz, 1H), 6.83–6.79 (m, 2H), 4.19 (q, $J = 10.8$ Hz, 1H), 4.17–4.11 (m, 1H), 4.09–4.05 (m, 2H), 3.74–3.70 (m, 1H), 3.29–3.25 (m, 1H), 3.10 (dd, $J = 16.1, 2.5$ Hz, 1H), 3.03–2.96 (m, 3H), 2.66–2.65 (m, 1H), 2.36–2.35 (m, 1H), 1.62–1.49 (m, 3H), 1.34–1.29 (m, 2H), 1.18 (t, $J = 7.1$ Hz, 3H), 0.91 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (175 MHz, CDCl_3) δ 171.6, 156.1, 140.3, 132.5, 129.3, 123.9, 121.8, 120.3, 61.0, 53.2, 51.9, 50.0, 38.1, 36.8, 30.6, 26.7, 20.4, 14.3; IR (KBr) ν 2925, 1730, 1628, 1460, 1423, 1366, 1351, 1307, 1239, 1176, 1106, 1029, 930, 853, 734 cm^{-1} ; HRMS (quadrupole, EI) m/z calcd for $\text{C}_{19}\text{H}_{26}\text{N}_2\text{O}_3$ $[\text{M}]^+$ 330.1943, found 330.1944.

General Procedure and Characterization for the Deprotection and Olefin Migration of 3f. To an oven-dried sealed tube charged with 7-allyl-*N*-butylindoline-1-carboxamide (**3f**) (51.6 mg, 0.2 mmol, 100 mol %) in EtOH (2 mL) was added aqueous *s*-KOH solution (1 mL) at room temperature. The reaction mixture was stirred for 20 h at 100 $^{\circ}\text{C}$ and cooled to 0 $^{\circ}\text{C}$. The reaction mixture was neutralized with aqueous *s*- NH_4Cl solution, and partitioned between EtOAc and H_2O . The organic layer was dried over MgSO_4 and concentrated in vacuo. The residue was purified by flash column

chromatography (silica gel, *n*-hexanes/EtOAc = 15:1) to afford 22.3 mg of **6c** in 70% yield.

(*E*)-7-(*Prop-1-enyl*)indoline (**6c**). Brown sticky solid; ¹H NMR (700 MHz, CDCl₃) δ 7.02 (d, *J* = 7.7 Hz, 1H), 6.97 (d, *J* = 7.2 Hz, 1H), 6.70 (t, *J* = 7.4 Hz, 1H), 6.34 (dd, *J* = 15.8, 1.6 Hz, 1H), 6.10 (dq, *J* = 15.7, 6.5 Hz, 1H), 3.57 (t, *J* = 8.4 Hz, 2H), 3.03 (t, *J* = 8.4 Hz, 1H), 1.86 (dd, *J* = 6.5, 1.7 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 147.7, 130.3, 127.6, 126.6, 125.2, 123.1, 121.2, 119.9, 47.3, 30.0, 19.1; IR (KBr) ν 3363, 2921, 2850, 1731, 1647, 1590, 1433, 1373, 1333, 1248, 1056, 962, 749 cm⁻¹; HRMS (quadrupole, EI) *m/z* calcd for C₁₁H₁₃N [M]⁺ 159.1048, found 159.1046.

General Procedure and Characterization for Oxidation of **6c**.

To an oven-dried sealed tube charged with **6c** (47.8 mg, 0.3 mmol, 100 mol %) in acetone (1 mL) was added 10% Pd/C (8.8 mg) at room temperature. The reaction mixture was stirred for 20 h at 80 °C and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc = 20:1) to afford 35.4 mg of **6d** in 75% yield.

(*E*)-7-(*Prop-1-enyl*)-1*H*-indole (**6d**). Yellow sticky solid; ¹H NMR (700 MHz, CDCl₃) δ 8.32 (br s, 1H), 7.53 (d, *J* = 7.8 Hz, 1H), 7.22–7.20 (m, 1H), 7.18 (d, *J* = 7.2 Hz, 1H), 7.09 (t, *J* = 7.6 Hz, 1H), 6.68 (dd, *J* = 15.7, 6.6 Hz, 1H), 6.57–6.56 (m, 1H), 6.32–6.27 (m, 1H), 1.98 (dd, *J* = 8.4 Hz, 1H), 1.86 (dd, *J* = 6.6, 1.7 Hz, 3H); ¹³C NMR (175 MHz, CDCl₃) δ 133.5, 128.3, 127.5, 127.0, 124.0, 121.9, 120.1, 119.9, 119.5, 103.1, 18.9; IR (KBr) ν 3376, 2922, 2848, 1593, 1434, 1248, 1195, 1056, 961, 748 cm⁻¹; HRMS (quadrupole, EI) *m/z* calcd for C₁₁H₁₁N [M]⁺ 157.0891, found 157.0891.

Experimental Procedure and Characterization of H/D Exchange Experiment without **2a**.

To an oven-dried sealed tube charged with *N*-butylindoline-1-carboxamide (**1f**) (65.5 mg, 0.3 mmol, 100 mol %), [RhCp*Cl₂]₂ (4.6 mg, 2.5 mol %), AgSbF₆ (10.3 mg, 10 mol %), and Cu(OAc)₂ (16.3 mg, 30 mol %) in THF (1 mL) was added MeOD (20 equiv). The reaction mixture was allowed to stir at room temperature for 20 h. The reaction mixture was diluted with EtOAc (3 mL) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc = 8:1) to afford 64.8 mg of **1f**/deuterio-**1f** (79% D incorporation) in 99% yield.

N-Butylindoline-1-carboxamide (**1f**) and *N*-Butyl-7-deuterioindoline-1-carboxamide (deuterio-**1f**). ¹H NMR (300 MHz, CDCl₃) δ 7.85 (d, *J* = 8.0 Hz, 0.21H), 7.12–7.07 (m, 2H), 6.85 (t, *J* = 7.3 Hz, 1H), 4.69 (br s, 1H), 3.83 (t, *J* = 8.5 Hz, 2H), 3.31–3.24 (m, 2H), 3.10 (t, *J* = 8.5 Hz, 2H), 1.56–1.47 (m, 2H), 1.40–1.28 (m, 2H), 0.92 (t, *J* = 7.2 Hz, 3H).

Experimental Procedure and Characterization of H/D Exchange Experiment with **2a**.

To an oven-dried sealed tube charged with *N*-butylindoline-1-carboxamide (**1f**) (65.5 mg, 0.3 mmol, 100 mol %), [RhCp*Cl₂]₂ (4.6 mg, 2.5 mol %), AgSbF₆ (10.3 mg, 10 mol %), and Cu(OAc)₂ (16.3 mg, 30 mol %) in THF (1 mL) were added allyl methyl carbonate (**2a**) (69.7 mg, 0.6 mmol, 200 mol %) and MeOD (20 equiv). The reaction mixture was allowed to stir at room temperature for 3 h. The reaction mixture was diluted with EtOAc (3 mL) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, *n*-hexanes/EtOAc = 8:1 to 6:1) to afford 57.3 mg of **3f** (74% yield) and 13.2 mg of **1f**/deuterio-**1f** (71% D incorporation, 20% yield), respectively.

Kinetic Isotope Effect (KIE) Experiment. To an oven-dried sealed tube charged with **1f** (32.8 mg, 0.15 mmol, 50 mol %), [RhCp*Cl₂]₂ (4.6 mg, 2.5 mol %), AgSbF₆ (10.3 mg, 10 mol %), and Cu(OAc)₂ (16.3 mg, 30 mol %) in *t*-AmOH (1 mL) were added allyl methyl carbonate (**2a**) (69.7 mg, 0.6 mmol, 200 mol %) and tetradecane (59.5 mg, 0.3 mmol) as an internal standard. In another reaction tube, deuterio-**1f** (32.9 mg, 0.15 mmol, 50 mol %, >99% D) was used instead of **1f**. The two reactions were allowed to stir at room temperature. An aliquot of each reaction mixture was taken at the time of 10, 20, 30, 40, and 50 min. The relative yield of each product was determined by GC–MS (tetradecane as an internal standard). A kinetic isotope effect value (*k_H/k_D*) of 2.89 was observed (see the Supporting Information for details).

■ ASSOCIATED CONTENT

📄 Supporting Information

Detailed kinetic isotope effect (KIE) experiment and ¹H NMR and ¹³C NMR copies of all compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Author

*Tel.: +82-31-290-7788. Fax: +82-31-292-8800. E-mail: insukim@skku.edu.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea Government (MSIP) (no. 2013R1A2A2A01005249).

■ REFERENCES

- (1) For selected reviews on C–H functionalization, see: (a) Ackermann, L. *Chem. Rev.* **2011**, *111*, 1315. (b) Cho, S. H.; Kim, J. Y.; Kwak, J.; Chang, S. *Chem. Soc. Rev.* **2011**, *40*, 5068. (c) Wencel-Delord, J.; Dröge, T.; Liu, F.; Glorius, F. *Chem. Soc. Rev.* **2011**, *40*, 4740. (d) Baudoin, O. *Chem. Soc. Rev.* **2011**, *40*, 4902. (e) McMurray, L.; O'Hara, F.; Gaunt, M. J. *Chem. Soc. Rev.* **2011**, *40*, 1885. (f) Bras, J. L.; Muzart, J. *Chem. Rev.* **2011**, *111*, 1170. (g) Kuhl, N.; Hopkinson, M. N.; Wencel-Delord, J.; Glorius, F. *Angew. Chem., Int. Ed.* **2012**, *51*, 10236. (h) Mousseau, J. J.; Charette, A. B. *Acc. Chem. Res.* **2013**, *46*, 412.
- (2) Oi, S.; Tanaka, Y.; Inoue, Y. *Organometallics* **2006**, *25*, 4773.
- (3) Tsai, A. S.; Brasse, M.; Bergman, R. G.; Ellman, J. A. *Org. Lett.* **2011**, *13*, 540.
- (4) Wang, H.; Schröder, N.; Glorius, F. *Angew. Chem., Int. Ed.* **2013**, *52*, 5386.
- (5) Feng, C.; Feng, D.; Loh, T.-P. *Org. Lett.* **2013**, *15*, 3670.
- (6) Fernández, I.; Hermatschweiler, R.; Breher, F.; Pregosin, P. S.; Veiros, L. F.; Calhorda, M. J. *Angew. Chem., Int. Ed.* **2006**, *45*, 6386.
- (7) (a) Yao, T.; Hirano, K.; Satoh, T.; Miura, M. *Angew. Chem., Int. Ed.* **2011**, *50*, 2990. (b) Makida, Y.; Ohmiya, H.; Sawamura, M. *Angew. Chem., Int. Ed.* **2012**, *51*, 4122. (c) Fan, S.; Chen, F.; Zhang, X. *Angew. Chem., Int. Ed.* **2011**, *50*, 5918. (d) Yu, Y. B.; Fan, S.; Zhang, X. *Chem.—Eur. J.* **2012**, *18*, 14643.
- (8) Zhang, Y. J.; Skucas, E.; Krische, M. J. *Org. Lett.* **2009**, *11*, 4248.
- (9) Zeng, R.; Fu, C.; Ma, S. *J. Am. Chem. Soc.* **2012**, *134*, 9597.
- (10) Ye, B.; Cramer, N. *J. Am. Chem. Soc.* **2013**, *135*, 636.
- (11) Yu, D.-G.; Gensch, T.; de Azambuja, F.; Vázquez-Céspedes, S.; Glorius, F. *J. Am. Chem. Soc.* **2014**, *136*, 17722.
- (12) Zhang, S.-S.; Wu, J.-Q.; Lao, Y.-X.; Liu, X.-G.; Liu, Y.; Lv, W.-X.; Tan, D.-H.; Zeng, Y.-F.; Wang, H. *Org. Lett.* **2014**, *16*, 6412.
- (13) (a) Joule, J. A.; Mills, K. *Heterocyclic Chemistry*; Blackwell Science Ltd.: Oxford, 2000. (b) Boger, D. L.; Boyce, C. W.; Garbaccio, R. M.; Goldberg, J. A. *Chem. Rev.* **1997**, *97*, 787. (c) Horton, D. A.; Bourne, G. T.; Smythe, M. L. *Chem. Rev.* **2003**, *103*, 893. (d) Chen, F.-E.; Huang, J. *Chem. Rev.* **2005**, *105*, 4671. (e) Trost, B. M.; Brennan, M. K. *Synthesis* **2009**, 3003.
- (14) Li, S.-M. *Nat. Prod. Rep.* **2010**, *27*, 57.
- (15) For selected examples on C3-allylations of indoles, see: (a) Butsugan, Y.; Nagai, K.; Nagaya, F.; Tabuchi, H.; Yamada, K.; Araki, S. *Bull. Chem. Soc. Jpn.* **1988**, *61*, 1707. (b) Zhu, X.; Ganesan, A. *J. Org. Chem.* **2002**, *67*, 2705. (c) McCubbin, J. A.; Hosseini, H.; Krokhin, O. V. *J. Org. Chem.* **2010**, *75*, 959. (d) Kimura, M.; Futamata, M.; Mukai, R.; Tamaru, Y. *J. Am. Chem. Soc.* **2005**, *127*, 4592. (e) Ma, S.; Yu, S.; Peng, Z.; Guo, H. *J. Org. Chem.* **2006**, *71*, 9865. (f) Zaitsev, A. B.; Gruber, S.; Pliiss, P. A.; Pregosin, P. S.; Veiros, L. F.; Wörle, M. *J. Am. Chem. Soc.* **2008**, *130*, 11604. (g) Sundararaju, B.; Achard, M.; Demerseman, B.; Toupet, L.; Sharma, G. V. M.; Bruneau, C. *Angew.*

Chem., Int. Ed. **2010**, *49*, 2782. (h) Montgomery, T. D.; Zhu, Y.; Kagawa, N.; Rawal, V. H. *Org. Lett.* **2013**, *15*, 1140.

(16) For selected examples on C2-allylations of indoles, see: (a) Gagnon, D.; Spino, C. *J. Org. Chem.* **2009**, *74*, 6035. (b) Yamakawa, T.; Ideue, E.; Shimokawa, J.; Fukuyama, T. *Angew. Chem., Int. Ed.* **2010**, *49*, 9262. (c) Auzzas, L.; Larsson, A.; Matera, R.; Baraldi, A.; Deschênes-Simard, B.; Giannini, G.; Cabri, W.; Battistuzzi, G.; Gallo, G.; Ciacci, A.; Vesce, L.; Pisano, C.; Hanessian, S. *J. Med. Chem.* **2010**, *53*, 8387. (d) Bennasar, M.; Solé, D.; Zulaica, E.; Alonso, S. *Org. Lett.* **2011**, *13*, 2042. (e) Li, B.; Ma, J.; Xie, W.; Song, H.; Xu, S.; Wang, B. *Chem.—Eur. J.* **2013**, *19*, 11863. (f) Kim, M.; Park, J.; Sharma, S.; Han, S.; Han, S. H.; Kwak, J. H.; Jung, Y. H.; Kim, I. S. *Org. Biomol. Chem.* **2013**, *11*, 7427.

(17) (a) Chakravarty, S.; Hart, B. P.; Jain, R. P. WO 2011103460 A1, 2011. (b) Kreft, A. F.; Caufield, C. E.; Failli, A. A.; Caggiano, T. J.; Greenfield, A. A.; Kubrak, D. M. US 5776967 A, 1998. (c) Reisch, J.; Adebajo, A. C.; Kumar, V.; Aladesanmi, A. J. *Phytochemistry* **1994**, *36*, 1073. (d) Meragelman, K. M.; McKee, T. C.; Boyd, M. R. *J. Nat. Prod.* **2000**, *63*, 427. (e) Wang, Y.; Gloer, J. B.; Scott, J. A.; Malloch, D. J. *Nat. Prod.* **1995**, *58*, 93.

(18) Chatani, N.; Yorimitsu, S.; Asaumi, T.; Kakiuchi, F.; Murai, S. *J. Org. Chem.* **2002**, *67*, 7557.

(19) (a) Kalyani, D.; Deprez, N. R.; Desai, L. V.; Sanford, M. S. *J. Am. Chem. Soc.* **2005**, *127*, 7330. (b) Shi, Z.; Li, B.; Wan, X.; Cheng, J.; Fang, Z.; Cao, B.; Qin, C.; Wang, Y. *Angew. Chem., Int. Ed.* **2007**, *46*, 5554. (c) Nishikata, T.; Abela, A. R.; Huang, S.; Lipshutz, B. H. *J. Am. Chem. Soc.* **2010**, *132*, 4978. (d) De, S.; Ghosh, S.; Bhunia, S.; Sheikh, J. A.; Bisai, A. *Org. Lett.* **2012**, *14*, 4466. (e) Jiao, L.-Y.; Oestreich, M. *Chem.—Eur. J.* **2013**, *19*, 10845. (f) Jiao, L.-Y.; Smirnov, P.; Oestreich, M. *Org. Lett.* **2014**, *16*, 6020.

(20) (a) Urones, B.; Arrayás, R. G.; Carretero, J. C. *Org. Lett.* **2013**, *15*, 1120. (b) Jiao, L.-Y.; Oestreich, M. *Org. Lett.* **2013**, *15*, 5374. (c) Song, Z.; Samanta, R.; Antonchick, A. P. *Org. Lett.* **2013**, *15*, 5662. (d) Pan, S.; Wakaki, T.; Ryu, N.; Shibata, T. *Chem.—Asian J.* **2014**, *9*, 1257.

(21) (a) Pan, S.; Ryu, S.; Shibata, T. *Adv. Synth. Catal.* **2014**, *356*, 929. (b) Ai, W.; Yang, X.; Wu, Y.; Wang, X.; Li, Y.; Yang, Y.; Zhou, B. *Chem.—Eur. J.* **2014**, *20*, 17653.

(22) Yang, X.-F.; Hu, X.-H.; Feng, C.; Loh, T.-P. *Chem. Commun.* **2015**, DOI: 10.1039/C4CC09330E.

(23) (a) Pan, C.; Abdokader, A.; Han, J.; Cheng, Y.; Zhu, C. *Chem.—Eur. J.* **2014**, *20*, 3606. (b) Shin, K.; Chang, S. *J. Org. Chem.* **2014**, *79*, 12197. (c) Hou, W.; Yang, Y.; Ai, W.; Wu, Y.; Wang, X.; Zhou, B.; Li, Y. *Eur. J. Org. Chem.* **2015**, DOI: 10.1002/ejoc.201403355.

(24) Kim, M.; Mishra, N. K.; Park, J.; Han, S.; Shin, Y.; Sharma, S.; Lee, Y.; Lee, E.-K.; Kwak, J. H.; Kim, I. S. *Chem. Commun.* **2014**, *50*, 14249.

(25) Kim, M.; Sharma, S.; Mishra, N. K.; Han, S.; Park, J.; Kim, M.; Shin, Y.; Kwak, J. H.; Han, S. H.; Kim, I. S. *Chem. Commun.* **2014**, *50*, 11303.

(26) For a recent review on formal SN-type reaction mechanism, see: Kuhl, N.; Schröder, N.; Glorius, F. *Adv. Synth. Catal.* **2014**, *356*, 1443.

(27) For the Rh(III)-catalyzed allylation using 2-vinylloxiranes, see: Yu, S.; Li, X. *Org. Lett.* **2014**, *16*, 1200.

(28) Simmons, E. M.; Hartwig, J. F. *Angew. Chem., Int. Ed.* **2012**, *51*, 3066.

(29) For the nucleophilic substitution mechanism under Rh(III) catalysis, see: (a) Yang, L.; Correia, C. A.; Li, C.-J. *Adv. Synth. Catal.* **2011**, *353*, 1269. (b) Grohmann, C.; Wang, H.; Glorius, F. *Org. Lett.* **2012**, *14*, 656. (c) Ng, K.-H.; Zhou, Z.; Yu, W.-Y. *Org. Lett.* **2012**, *14*, 272.